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At the time of writing I am compiling a list of common names for the British species in the Order. Historically, only two have such names, *Chelifer cancroides*, the House Scorpion and *Cheiridium museorum*, the Book Scorpions. Generally speaking I am against the use of such names as it can cause confusion and also make it more difficult — twice as many names to remember. However, I do see the argument for having them in that none-specialist, such as planners and developers feel easier when they are used. They also easier to pronounce, at least generally (some common names use bastardised Latin names) which makes them less off putting for the lay public and non-specialists.

The following is a list of suggested names only, they have yet to be commented on.

Binomial

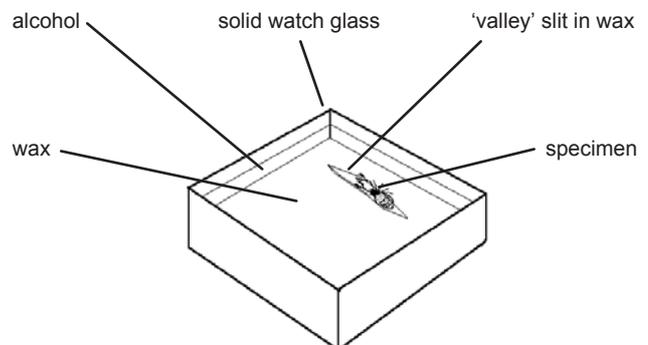
- Chthonius halberti*
- Chthonius tetrachelatus*
- Chthonius kewi*
- Chthonius ischnocheles*
- Chthonius tenuis*
- Chthonius orthodactylus*
- Neobisium carcinoides*
- Neobisium maritimum*
- Neobisium carpenteri*
- Roncus lubricus*
- Roncocreagris cambridgei*
- Cheiridium museorum*
- Chernes cimicoides*
- Dendrochernes cyrneus*
- Dinocheirus panzeri*
- Lamprochernes nodosus*
- Lamprochernes savignyi*
- Lamprochernes chyzeri*
- Allochernes powelli*
- Allochernes wideri*
- Pselaphochernes dubius*
- Pselaphochernes scorpioides*
- Chelifer cancroides*
- Dactylochelifer latreillei*
- Larca lata*
- Microbisium brevifemoratum*
- Withius piger*

Common Name

- Halbert's chthonid
- Dimple-clawed chthonid
- Kew's chthonid
- Common chthonid
- Dark-clawed chthonid
- Straight-fingered chthonid
- Moss neobisid
- Shore neobisid
- Carpenter's neobisid
- Reddish two-eye chelifer
- Cambridge's two-eye chelifer
- Book scorpion
- Common tree-cherne
- Large tree-cherne
- Terrible-clawed chernes
- Knotty shining claw
- Savigny's shining claw
- Chyzer's shining claw
- Powell's chernes
- Wider's tree-cherne
- Small chernes
- Compost chernes
- House scorpion
- Marram grass chelifer
- Oak-tree chelifer
- Bog chelifer
- Lazy chelifer

Records Received

The number of records received since the last *Newsletter* has been about the same as the previous year. Thanks again to all those who have contributed - you know who you are! To these and other readers of this *Newsletter* please keep the records coming — even what you consider as 'repeat' records are welcome. Such records, over time, can help the understanding of change in a habitat or geographical area. If in doubt send the specimens and they will be checked and returned.



Viewing Difficult Characters

Viewing some characters can be a problem as the specimen will not cooperate and 'sit' in the right position. One way round this is to cut off the pedipalp or what ever and tilt it in such a way that the features you are trying to see are visible. This of course is fiddly and also damages the specimen — it is easy to loose the amputated appendage. By filling a watch glass with glass or resin beads (as from some water purifiers) a specimen can be tilted and holds its position through the action of the substrate. Another technique, which I commonly employ, is to use a wax filled solid watch glass or re-cycled cover-slip plastic box. Paraffin (candle) wax

or bees' wax is melted into the container to a depth of a couple of millimetres or so. A fine scalpel or similar blade is then used to cut a fine trench about 4 to 5 mm long, the depth tapering towards one end. By carefully tilting a specimen it is easy to wedge it in the trench at virtually any angle and thus make it possible to view the feature under scrutiny.

Collecting Tips

Many of you will be familiar with a variety of collecting techniques, but others may not be so conversant. Pseudoscorpions being small and secretive can present problems when it comes to finding them. A lot are found and collected not as part of deliberate search for the beasts, but by accident whilst looking for other arthropods. As a result the methods listed below include techniques which are primarily aimed at, for example, beetles or flies. However, the Coleopterist and Dipterist can be important sources of pseudoscorpion data even if they only consider their appearance as an oddity. Here are a few ideas to consider.

1. Hand sorting.

This is the most commonly employed technique. It is laborious and not particularly effective, nevertheless if you are on holiday it is straight forward and easy to do. A tray, preferably white, is all that is needed. Some sun shine helps too! Scatter a handful of debris (leaf litter, rotting wood, barn debris, seaweed, old bird nest [bear in mind the Wild Life & Countryside Act and amendments] or what ever, over the tray, wait and watch, poking about occasionally. Adults are the most likely to be spotted. *Neobisium carcinoides* tends to move slowly about unlike *Chthonius ischnocheles* which can move slowly, but often has a darting gait – either backwards or forwards. A pooter (aspirator) or fine paint spittle moistened paint brush can be used to pick individuals up. Place them in either a vial containing a few pieces of fresh grass (to keep the humidity up and prevent desiccation) if you want them alive. Drop into 70% alcohol if you want to keep them for further study.

2. Bark prising.

Again – be considerate and conservation minded. Don't go 'destroying tree habitats hither and thither else you could be in trouble, and it is very antisocial. Look at the surface of the timber exposed then the inner surface of the piece of bark that has been removed. Poke about with a knife and aspirate or use a paint brush to collect individuals. Look out for silken chambers too which can resemble the silken 'nests' of some small spiders. On removing the bark silken chambers are often torn leaving the pseudoscorpion inside exposed inside a decapitated igloo.

3. Sieving

Really part of hand sorting, but using a sieve to restrict the particles fallen on to a white tray. Hand and mechanical sieves (shakers) can be used – the domestic garden sieve is perfectly adequate and works well.

4. Turning over objects

Drift wood on the strand line, logs, bags in barns and rocks can all yield specimens when turned over. DON'T forget though, put the log or what ever back again as you found it.

5. Bag a tree

Rather than scrub around inside a rotten tree or prise its bark off, tie an onion bag full of wood wool or shavings (try synthetic packaging) around a tree and leave for 'x' weeks. Collect later and sort or extract (see later) the arthropods that have taken refuge in your artificial debris.

6. Band a tree

Arthropods, including pseudoscorpions can be trapped, again with no damage to the tree, using bands of corrugated material, bubble wrap etc. tied around a tree and left for some weeks.

7. Buried tray

Shingle, pebbles and loose shale is difficult to search through. Try burying a tray or trays. Ideally use the solid kind which bakeries use with a few holes drilled in to allow for drainage. Scrape a hole big enough to bury the tray to a depth about twice as deep as the tray and fill with shingle. Leave a week and then carefully remove and carefully remove and sort through the pebbles.

8. False rocks

Rock crevices are notoriously difficult to sample. Doing so can be extremely destructive and antisocial too! Why not do what one enterprising student did. Make up 'LEGO' into blocks and wedge them into crevices on the shore. Leave for a week or more then carefully remove and place in a bag. Back at home or in more comfort, dismantle the block(s) and collect the creatures hiding therein.

9. Pit falls

An old favourite, but they work and may indicate the surface activity of certain species. There are all kinds of ways of doing this. One way is to use plastic drink cups dug into the ground, covered to prevent the ingress of rain and containing 30ml or so of 10% antifreeze to act as a temporary preservative (not fixative). Inspect regularly.

10. Deep soil pit falls

Special deep soil, hypogean traps can be made from a couple of drinks bottles. To date they have caught several rare and unusual beetles as well as the occasional pseudoscorpion.

11. Light traps

Although designed for moths and other nocturnal creatures, pseudoscorpions can be caught too. Not flying of course, but phoretic on the bodies and limbs of beetles and other insects. Even... see article... can be found

12. Malaise traps

These can also yield specimens which have hitched a lift on some unsuspecting insect.

13. Tullgren Funnel

This is one of the most efficient ways of 'extracting' arthropods, including pseudoscorpions. It relies on the creatures following the humidity gradient they most like. By applying warmth to the top surface of the sample a humidity gradient is set up in the funnel as the sample slowly dries. With progressive desiccation the gradient shifts downwards as do the arthropods which eventually fall into the collecting pot. A simple device can be made from a coarse (3-5mm mesh) sieve over a large plastic funnel above a collecting vessel. Heat using a 25 Watt bulb in a lamp enclosed above the sample.

There are, no doubt other techniques and tricks to find and capture specimens. Only a few are given here. If you have your own particular methods let me know.